

Article ID: 199695
DOI: 10.5586/aa/199695

Publication History

Received: 2024-08-30
Accepted: 2025-01-01
Published: 2025-06-26

Handling Editor

Ewa Muszyńska-Sadłowska;
Department of Botany,
Institute of Biology, Warsaw
University of Life Sciences,
Poland, <https://orcid.org/0000-0003-0419-4597>

Authors' Contributions

NMAA, SMA: Research concept and design; NMAA, HMA, AAA-Z: Collection and/or assembly of data; SMA, HMA, AAA-Z: Data analysis and interpretation; HMA, AAA-Z: Writing the article; NMAA: Critical revision of the article; NMAA, SMA, HMA, AAA-Z: Final approval of the article

Funding

The research was funded by the Deanship of Scientific Research at Umm Al-Qura University for supporting this work by Grant Code: (22UQU4280488DSR02).

Competing Interests

No competing interests have been declared.

Copyright Notice

© The Author(s) 2025. This is an open access article distributed under the terms of the [Creative Commons Attribution License](#), which permits redistribution, commercial and noncommercial, provided that the article is properly cited.

ORIGINAL RESEARCH

Synergistic Effects of Titanium Dioxide Nanoparticles and Ascorbic Acid on Drought Tolerance in Coffee Plants through Osmoprotection and Antioxidant Defense

Nadiyah M. Alabdallah^{1,2,*}, Saleh M. Alluqmani³, Hana Mohammed Almarri^{2,4}, Asla A. AL-Zahrani^{2,5}

¹ Department of Biology, College of Science, Imam Abdulrahman Bin Faisal University, P.O. Box 1982, 31441, Dammam, Saudi Arabia

² Basic and Applied Scientific Research Center – College of Science – Imam Abdulrahman Bin Faisal Abdulrahman bin Faisal University, P.O. Box 1982, Dammam, 31441, Saudi Arabia

³ Department of Physics, Faculty of Applied Science, Umm Al-Qura University, Makkah 21955, Saudi Arabia

⁴ Department of Physics, College of Science, Imam Abdulrahman Bin Faisal University, P.O. Box 1982, Dammam, 31441, Saudi Arabia

⁵ Department of Chemistry, College of Science, Imam Abdulrahman Bin Faisal University, P.O. Box 1982, Dammam, 31441, Saudi Arabia

*To whom correspondence should be addressed. Email: nmalabdallah@iau.edu.sa

Abstract

Titanium dioxide nanoparticles (TiO₂NPs) and ascorbic acid (AsA) have been shown to enhance plant productivity. However, the effects of applying a combined dose of these substances to plants under drought stress are not well understood. Therefore, this study aimed to assess the effectiveness of applying TiO₂NPs (10 ppm) and AsA (50 ppm) to the leaves of coffee plants (*Coffea arabica* cv. Khawlani) subjected to drought conditions at 50% and 20% of their field capacity (FC). The study's findings demonstrated that coffee plants subjected to drought stress exhibited reductions in root and shoot length, as well as decreases in fresh and dry weights, relative water content (RWC), while also showing increases in proline levels, total soluble carbohydrates (TSC), malonaldehyde (MDA) content, and antioxidant activities (superoxide dismutase SOD and catalase CAT activity). However, in drought stress conditions at 50% and 20% FC, TiO₂NPs enhanced shoot length by 23%, 12%, and 33%, while root length improved by 26%, 12%, and 38%. AsA treatments further increased shoot length by 44%, 39%, and 51%, along with root growth of 45%, 16%, and 54%, respectively. Following the application of TiO₂NPs and AsA, these drought-stressed (50% and 20% FC) coffee plants displayed enhanced fresh and dry weights, increased RWC, and higher activities of SOD and CAT, along with decreased levels of MDA, and so on. Taken together, TiO₂NPs and AsA, whether applied individually or in combination, could serve as highly effective components for promoting the development of coffee plants by enhancing their physiological functions.

Keywords

carbohydrates; field capacity; growth; proline; malonaldehyde

1. Introduction

Drought limits plant growth and development, posing a significant challenge to crop productivity (Bakır et al., 2022). Under drought stress, crop yields are adversely affected (Hasan et al., 2023), and the production of harmful reactive oxygen species (ROS) escalates rapidly. If not efficiently neutralized by the plant antioxidant defense mechanisms, these ROS can lead to extensive cellular damage (Hasan et al., 2021a;

Alabdallah et al., 2021). This damage includes harm to cellular membranes and components in plants (Jahan et al., 2022; Hasan et al., 2021b). For example, during drought stress, elevated levels of MDA are often utilized as markers of oxidative damage to the plant cell (Khan et al., 2017; Abdelaal et al., 2022). Moreover, plants have evolved antioxidant defense mechanisms, particularly by enhancing the activities of SOD and CAT enzymes, to control ROS levels during drought stress (Rahman et al., 2022; Contreras-Oliva et al., 2022).

Coffee is a widely enjoyed beverage, ranking as the second most traded commodity worldwide (Contreras-Oliva et al., 2022). However, coffee yields have come under threat due to an escalation in both the frequency and severity of drought conditions (Silva et al., 2022). This situation poses potential risks not only to the quantity of coffee produced but also to its quality (DaMatta et al., 2018). Drought is a significant environmental challenge, and its adverse effects have been substantial, resulting in an alarming 80% reduction in global coffee production (Merga et al., 2023). Therefore, it is important to formulate strategies aimed at bolstering coffee's resilience to drought and enhancing its overall productivity.

The application of titanium dioxide nanoparticles (TiO₂NPs) and ascorbic acid (AsA) has demonstrated the ability to enhance the drought stress tolerance in plants (Celi et al., 2023; Karvar et al., 2022). Titanium dioxide (TiO₂), found in everyday items like plastics and cosmetics, poses environmental risks, potentially leading to human health concerns (Menard et al., 2011). Toxicity is a major issue in the environment, directly related to various factors, including chemical compounds (Melo et al., 2022). Human exposure to TiO₂ can occur through various routes, such as consuming contaminated water from rivers where individuals have used sunscreen. In agriculture, TiO₂ in pesticides and fertilizers enhances plant growth and soil fertility, with its photocatalytic properties aiding in pesticide breakdown (Menard et al., 2011). Its agricultural use may result in widespread environmental distribution, posing risks through contaminated food crops or fish (Jovanovc, 2015). However, TiO₂NPs have garnered significant attention due to their excellent compatibility with living organisms, low toxicity, strong capacity to neutralize harmful free radicals, and high availability for biological processes (Wang et al., 2022). TiO₂NPs are considered to have the potential to enhance crop productivity (Li et al., 2018). For instance, TiO₂NPs with melatonin increase the rate of photosynthesis, the activity of rubisco, the potential of antioxidant enzymes, and the production of chlorophyll in plants (Sheikhalipour et al., 2023). Conversely, AsA serves as a crucial growth regulator that safeguards plant tissues from the harmful impacts of oxygen radicals (Celi et al., 2023). For instance, AsA acts as a co-factor for enzymes responsible for hormone production and the regeneration of enzymatic antioxidants in plants (Castro et al., 2023).

However, despite these advancements, there has been limited research into the combined use of ascorbic acid and TiO₂NPs in coffee plants facing drought stress. Therefore, it is urgent to explore these novel findings by investigating the impact of ascorbic acid and TiO₂NPs on coffee plants. This has led to the hypothesis that applying ascorbic acid along with TiO₂NPs could be a valuable approach to enhance the drought tolerance of coffee plants. Consequently, our objective was to assess the following: (i) The effectiveness of ascorbic acid and TiO₂NPs in mitigating the detrimental effects of drought stress on coffee plant growth. (ii) The influence of ascorbic acid and TiO₂NPs on physicochemical characteristics that contribute to improved crop yield. (iii) The possibility to cultivate coffee plants with reduced water consumption without experiencing a significant reduction in yield.

2. Materials and methods

2.1. Growth conditions

This study was designed to evaluate the effect of TiO₂NPs and AsA on the drought tolerance of coffee plants (*Coffea arabica* cv. Khawlani) in controlled conditions. The experiments were conducted in a controlled greenhouse at the Department of Biology, Imam Abdulrahman Bin Faisal University, while the biochemical analysis was performed at Umm Al-Qura University, Saudi Arabia. Coffee seeds used in the study

were sourced from the Jazan area in Saudi Arabia (latitude 16.8894°N, longitude 42.5706°E). To prepare the coffee seeds for planting, they were first sterilized with a 1% sodium hypochlorite (NaOCl) solution for 10 minutes and then rinsed with tap water for an additional minute. The treated seeds were then transferred into plastic pots, each filled with a mixture of garden soil.

The greenhouse maintained day/night temperatures of 22°C/16°C and relative humidity in the range of 60–70%. The plants were watered twice a week to reach a soil moisture level corresponding to 100% field capacity (FC), which served as the control condition. Drought stress was induced by adjusting watering to achieve 50% FC and 20% FC, simulating moderate and severe drought conditions, respectively, for two weeks. TiO₂NPs and AsA were applied through foliar spray using a hand-held sprayer to ensure even distribution. The spray was applied starting in the 25th up to 30th weeks during the drought period, with concentrations of 10 ppm for TiO₂NPs and 50 ppm for AsA, combining 10 ppm TiO₂NPs and 50 ppm AsA for each application per plant. Each plant received a total spray volume of 30 mL per application, applied twice a week during the drought period. The coffee plants representing a mature growth stage of seedlings were harvested at 30 weeks for physiological and biochemical analyses.

2.2. TiO₂NP preparations and experimental treatments

We synthesized TiO₂NPs using sol-gel techniques. Titanium tetraisopropoxide (TIP), with a purity level of 99.9%, was combined with acetic acid (purity ≥ 99.7%) and ethanol (purity ≥ 99.9%) at ambient room temperature. The mixture was prepared in a specific molar ratio of 0.5:0.05:4.5 for TIP, acetic acid, and ethanol, respectively. The solution was then continuously stirred for 1.5 hours to ensure proper blending and homogenization. Following this, the resulting mixture was subjected to drying in an oven set at 100°C, where it was allowed to dry for 12 hours. Subsequently, the dried material underwent thermal treatment, reaching temperatures in the range of 500 to 550°C in a furnace for 2 hours, with a heating rate of approximately 0.16°C per minute. The size and shape of the heated samples were characterized as spherical with dimensions ranging from 30 to 55 nm using a scanning electron microscope (SEM). In this experiment, we employed four different treatments: the control group, 10 ppm TiO₂NPs, 50 ppm AsA, and a combination of 10 ppm TiO₂NPs with 50 ppm AsA.

2.3. Measurements of growth parameters

Coffee plant shoot and root lengths were measured using the metric system, and the units were reported in centimeters (cm). On the days of harvest, length-related attributes were determined using a measuring tape after the plants were gently uprooted. The roots and shoots were rinsed with tap water, and their lengths were measured in centimeters (cm). The weight of fresh coffee plant shoots and roots were determined using a digital balance (HR–200). The dry weights of the shoots and roots were recorded after they had been dried at 60°C.

2.4. Relative water content (RWC)

The RWC in the coffee plants was measured following the protocol described by Barrs and Weatherley (1962). It was determined using the following formula: $RWC (\%) = [(Fresh\ weight, FW - Dry\ weight, DW) / (Turgid\ weight, TW - Dry\ weight, DW)] \times 100$. To obtain the TW, the leaves were submerged in distilled water for 6 hours, allowing them to reach full turgidity. The DW was measured after the leaves were thoroughly dried to remove all moisture content.

2.5. MDA content measurements

The MDA concentration in the sample was quantified by following the protocol established by Heath and Packer (1968). To begin, 0.5 g of coffee fresh leaf tissue were ground and homogenized with 5 mL of a 10% trichloroacetic acid (TCA) solution. The homogenate was then subjected to centrifugation at 15,000×g for 10 minutes to

separate the solid and liquid phases. After centrifugation, 2 mL of the supernatant were carefully collected and combined with 4 milliliters of a 0.6% thiobarbituric acid (TBA) solution. The resulting mixture was thoroughly blended to ensure uniformity. The mixture was subsequently heated to 100°C for 15 minutes to promote the reaction between MDA and TBA, which produces a colored complex. After heating, the reaction was immediately stopped by rapidly cooling the mixture on ice. The absorbance of this complex was measured at 532 nm to quantify the MDA concentration, as this wavelength is specific for the MDA–TBA complex. In addition, absorbance at 600 nm was measured to correct for any non-specific turbidity and light scattering in the sample, ensuring accuracy. The extinction coefficient used for the MDA–TBA complex was 155 mM⁻¹ cm⁻¹.

2.6. Proline content determination

Proline content was determined by the protocol of Bates et al. (1973). 0.2 g of fresh leaf tissue was first homogenized with 10 mL of a 3% (w/v) sulfosalicylic acid solution. The homogenate was then centrifuged at 15,000×g for 10 minutes to separate the solid and liquid phases. After centrifugation, 2 mL of the clear supernatant were carefully transferred and mixed with 2 mL of glacial acetic acid and 2 mL of an acid ninhydrin solution in test tubes. The tubes were then placed in a boiling water bath for 1 hour to allow the reaction to take place, which facilitated the formation of a chromophore. Following the heating step, 4 mL of toluene were added to extract the reaction product into the organic phase. The absorbance of the toluene phase containing the chromophore was measured at 520 nm.

2.7. Determination of total soluble carbohydrates (TSC)

TSC were determined following the protocol established by Dubois et al. (1956). In this procedure, anthrone was prepared by dissolving it in 45 mL of 95.5% (v/v) sulfuric acid. For the reaction, a mixture was created by combining 50 µL of the alcoholic extract with 950 µL of deionized water, resulting in a total volume of 1000 µL. To this reaction mixture, 2000 µL of the prepared anthrone solution were gradually added while keeping the reaction vessel in an ice bath to prevent overheating. After the addition, the mixture was incubated in a water bath at 100°C for 3 minutes to promote the reaction between anthrone and the carbohydrates in the extract. Once the incubation was complete, the mixture was allowed to cool to room temperature naturally. Glucose was used as the reference standard for this procedure. The absorbance of the resulting solution was measured at 620 nm using a spectrophotometer to quantify the carbohydrate content in the sample.

2.8. Antioxidant activity measurements

A fresh coffee leaf sample weighing 0.3 g was placed in 5 mL of phosphate buffer, which had a concentration of 50 mmol/L and a pH of 7.0. To enhance the extraction efficiency, 10 g/L of polyvinylpyrrolidone (PVP) was added. The mixture was then subjected to centrifugation at 15,000×g for 20 minutes to separate the solid plant material from the liquid extract. After centrifugation, the supernatant was carefully collected and used for the preliminary analysis of enzyme activities.

The activity of SOD was determined based on the method described by Hasan et al. (2021b), with slight modifications. The reaction mixture for SOD activity included 0.1 mL of 60 µM riboflavin, 1.5 mL of 26 µM L-methionine, 1.0 mL of 225 µM nitroblue tetrazolium (NBT), 0.75 mL of 1% Triton-X, and 15 µL of 2 mM EDTA, all dissolved in 27.0 mL of sodium-potassium phosphate (NaKPi) buffer at pH 7.8. To this reaction mixture, 0.5 mL of the enzyme extract was added to make a total volume of 3.0 mL. The tubes containing the mixture were incubated at 25°C for 10 minutes, shielded from direct light. The reaction was initiated by exposing the tubes to light from a 15-watt fluorescent lamp (pre-heated, 6500 K). The reaction was allowed to proceed for 20 minutes under light and the SOD activity was quantified by measuring the absorbance of the reaction at 560 nm, which reflects the enzyme ability to inhibit the photochemical reduction of NBT.

CAT activity was evaluated following the method outlined by Aebi (1984). In this procedure, 20 µL of the enzyme extract was mixed with 830 µL of potassium phosphate buffer (50 mM, pH 7.0). The enzymatic reaction was initiated by adding 150 µL of 0.05M hydrogen peroxide (H₂O₂). The reaction progress was monitored by measuring the decrease in absorbance at 240 nm over one minute, reflecting the CAT activity as it breaks down H₂O₂ into water and oxygen.

2.9. Statistical analysis

In this study, a Completely Randomized Design (CRD) was implemented for the treatment application. A one-way analysis of variance (ANOVA) was conducted to evaluate the data, ensuring that any differences between groups were statistically analyzed. To further explore and compare the specific group differences, Fisher's Least Significant Difference (LSD) was employed, with a significance level set at $p \leq 0.05$. All data processing and statistical calculations were carried out using Minitab-17 software.

3. Results

Scanning electron microscopy (SEM) analysis was carried out to verify the successful synthesis of TiO₂NPs and provided detailed insights into the morphology, size, and surface structure of the nanoparticles, confirming that the desired characteristics were achieved. After validation through SEM imaging, as illustrated in Figure 1, the synthesized TiO₂NPs were incorporated into experimental studies aimed at exploring their effectiveness in reducing the adverse effects of drought stress on coffee plants.

The growth of the coffee plants was observed to decrease significantly ($p \leq 0.05$) when subjected to the drought stress conditions, specifically at 50% and 20% field capacity (FC). For instance, all morphological parameters of the coffee plants exhibited a notable decline ($p \leq 0.05$) in response to the drought stress treatment (50% and 20% FC) (Figure 2, 3).

However, the diverse concentrations of TiO₂NPs and AsA treatments (10 ppm TiO₂NPs, 50 ppm AsA, and 10 ppm TiO₂NPs + 50 ppm AsA) led to varying degrees of improvement in these growth parameters (Figure 2, 3).

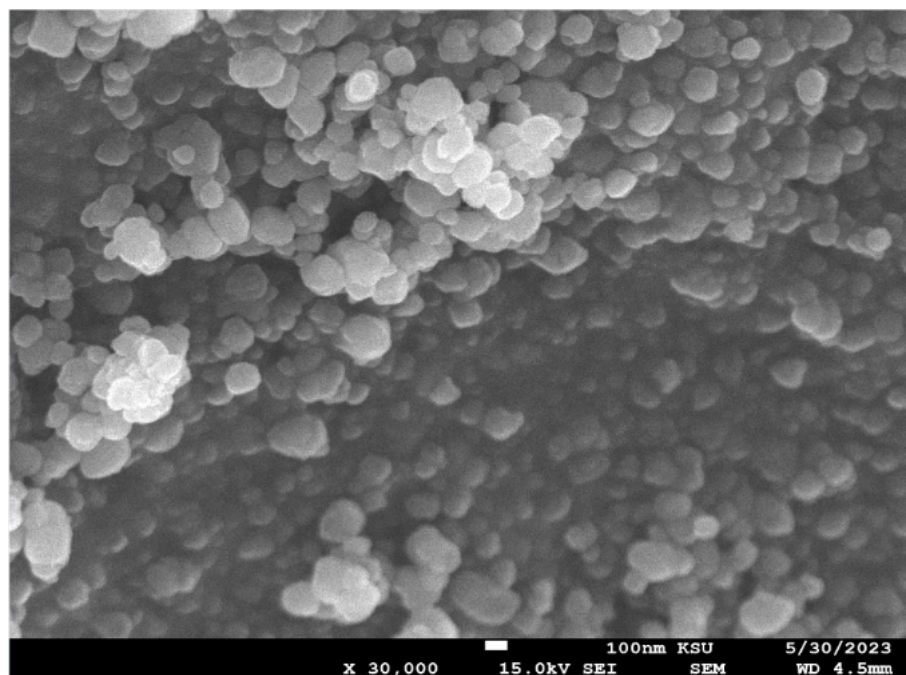


Figure 1 SEM techniques visualize the successful synthesis of TiO₂NPs by providing high-resolution imaging.

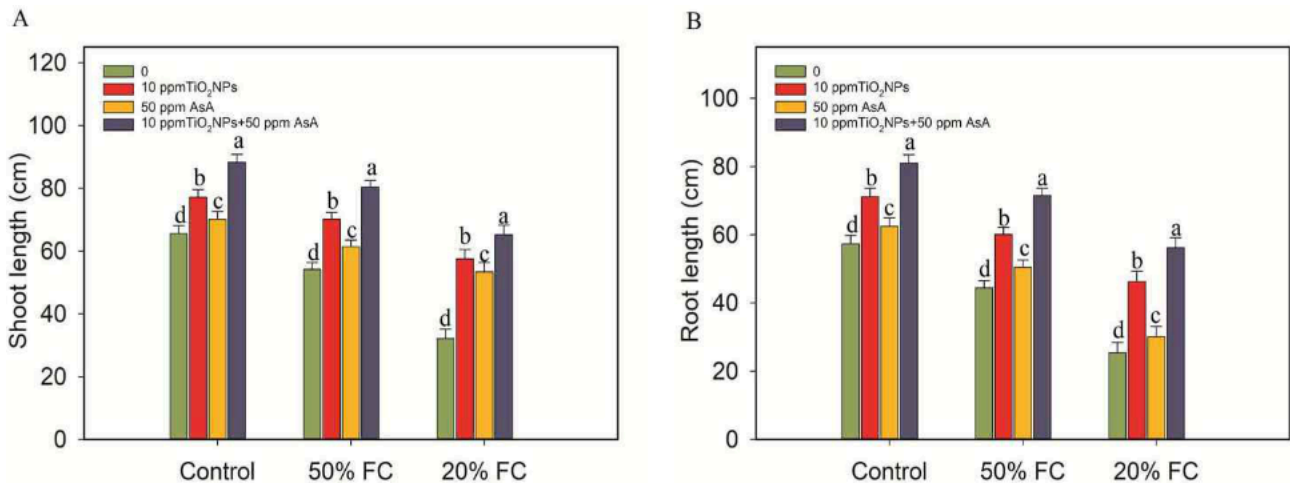


Figure 2 Impacts of TiO₂NPs and AsA on the shoot and root lengths of coffee plants during drought stress conditions (50% FC and 20% FC). Statistically significant differences at the $p \leq 0.05$ level are indicated by bars with different letter annotations.

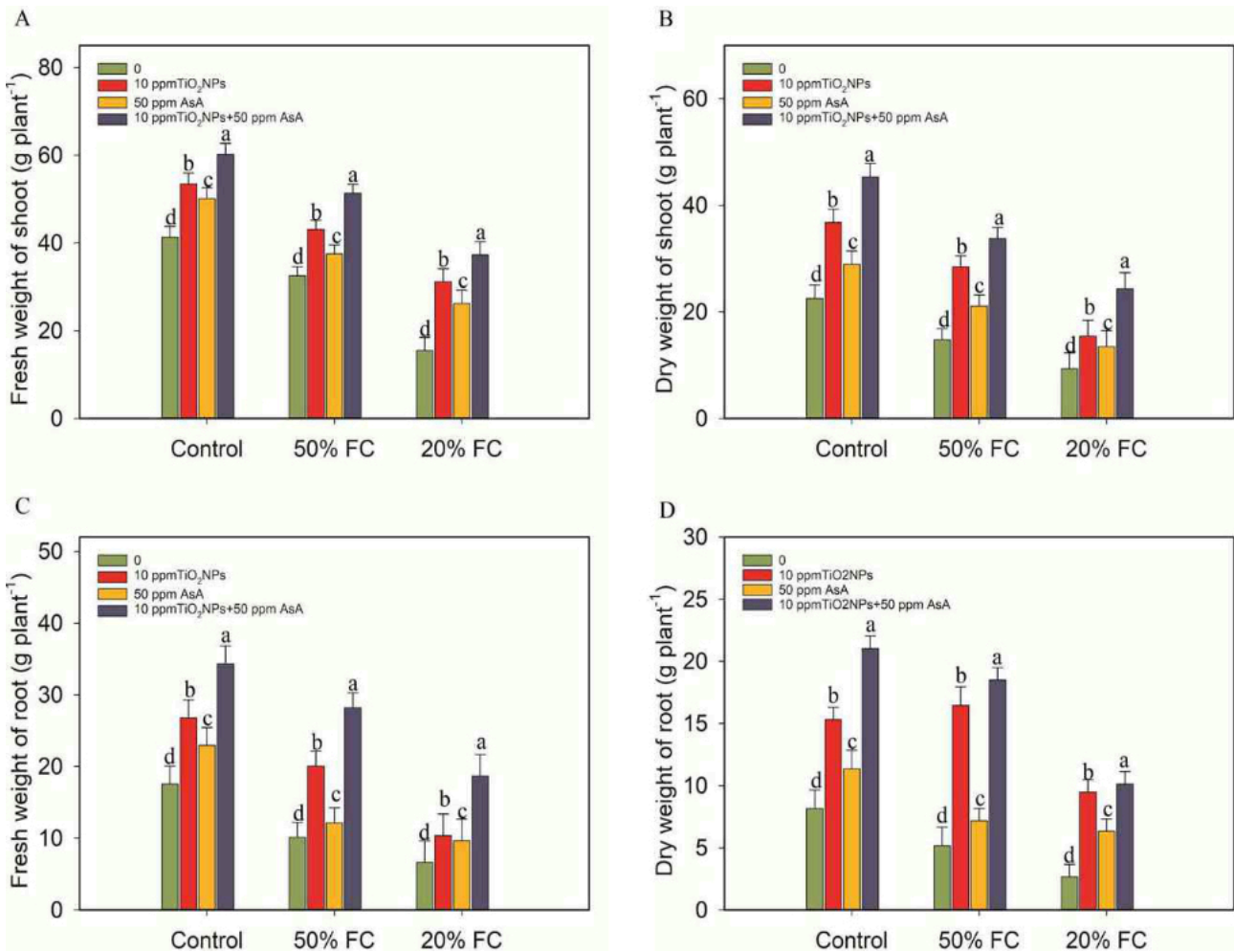


Figure 3 Impacts of TiO₂NPs and AsA on the fresh weight of shoot and root and dry weight of shoot and root of coffee plants during drought stress conditions (50% FC and 20% FC). Statistically significant differences at the $p \leq 0.05$ level are indicated by bars with different letter annotations.

The application of TiO₂NPs and AsA treatments (10 ppm TiO₂NPs, 50 ppm AsA, and 10 ppm TiO₂NPs + 50 ppm AsA) resulted in increased RWC by 8%, 6%, and 13%, respectively, when compared to the control.

In contrast, the drought treatments at 50% and 20% FC resulted in a substantial reduction in the RWC in the coffee plants, with decreases of 25% and 40%, respectively, when compared to the control group. However, the application of the TiO_2 NPs and AsA treatments led to notable improvements in RWC. Specifically, TiO_2 NPs enhanced RWC by 13%, 8%, and 18%, while the AsA treatments increased it by 15%, 10%, and 19% in coffee plants subjected to drought stress at 50% and 20% FC, respectively. Furthermore, drought stress significantly ($p \leq 0.05$) elevated the concentrations of organic compounds such as proline (Figure 4) and TSC (Figure 5) in the coffee plants.

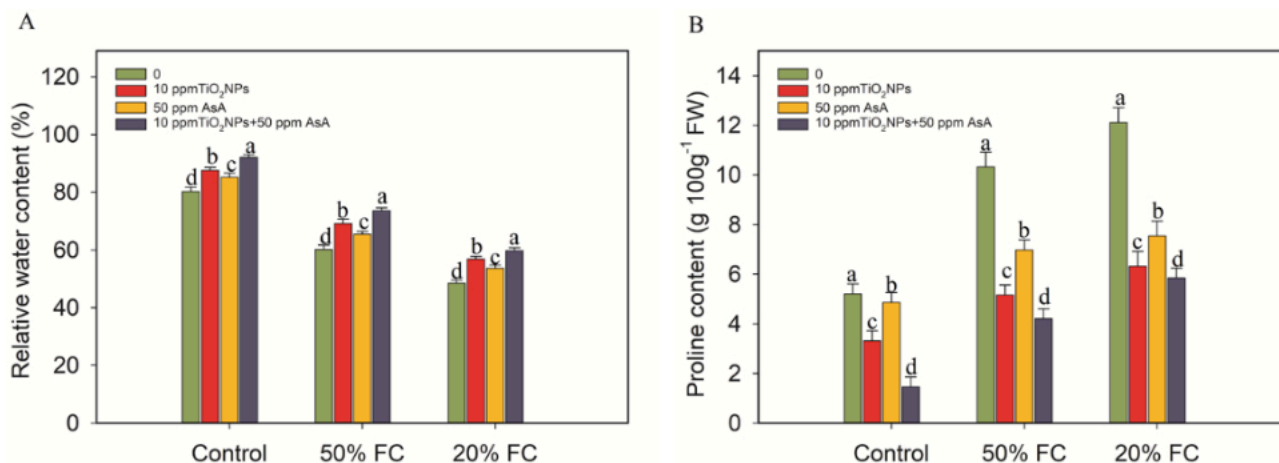


Figure 4 Impacts of TiO_2 NPs and AsA on the relative water and proline content in coffee plants during drought stress conditions (50% FC and 20% FC). Statistically significant differences at the $p \leq 0.05$ level are indicated by bars with different letter annotations.

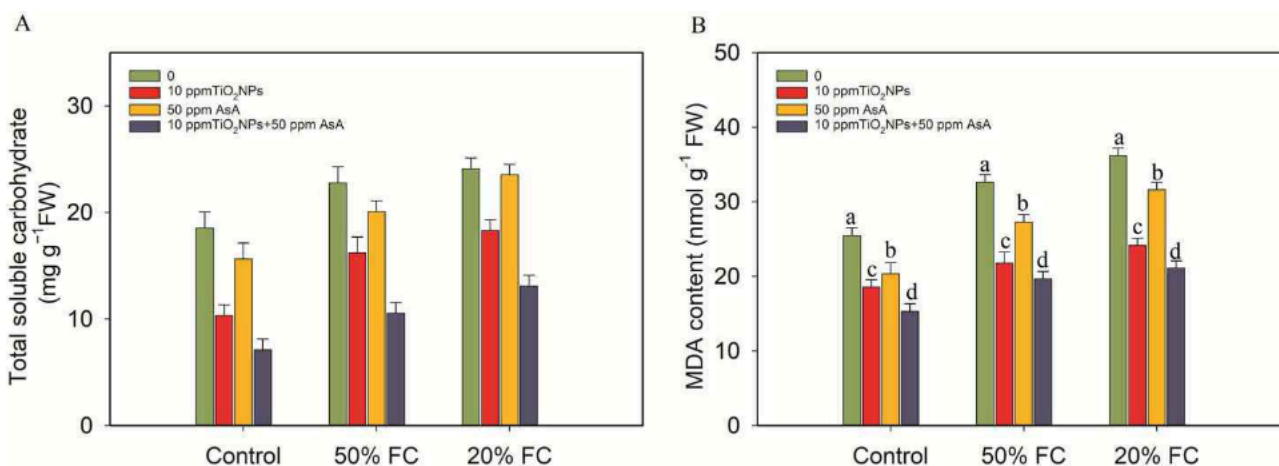


Figure 5 Impacts of TiO_2 NPs and AsA on the total soluble carbohydrates and MDA content in coffee plants during drought stress conditions (50% FC and 20% FC). Statistically significant differences at the $p \leq 0.05$ level are indicated by bars with different letter annotations.

In contrast, when the coffee plants were treated with TiO_2 NPs and AsA, a notable reduction in proline content was observed. Specifically, the TiO_2 NP treatments led to a decrease in proline levels by 50%, 32%, and 59%, while the AsA treatments resulted in reductions of 48%, 38%, and 52%, compared to the proline levels in the drought-stressed coffee plants at 50% and 20% FC. Similarly, the levels of TSC also dropped significantly with the application of TiO_2 NPs and AsA. TiO_2 NPs reduced TSC by 29%, 12%, and 54%, whereas the AsA treatments led to reductions of 24%, 2%, and 45%, respectively, when compared to the TSC levels in the drought-stressed plants at 50% and 20% FC (Figure 5).

In the drought stress conditions at 50% and 20% FC, there was a substantial increase of 22% and 29%, respectively, in the malondialdehyde (MDA) levels, compared to the control. However, the application of the TiO₂NPs and AsA treatments (10 ppm TiO₂NPs, 50 ppm AsA, 10 ppm TiO₂NPs + 50 ppm AsA) resulted in reductions of 33%, 16%, 40%, and 33%, 13%, 42%, respectively, in the MDA levels, compared to the coffee plants under drought stress (Figure 5).

The drought treatments at 50% and 20% FC caused a marked rise in the activities of SOD and CAT in the coffee plants. SOD activity increased by 77% and 89%, while CAT activity showed increases of 55% and 67%, respectively, when compared to the control plants. These results suggest that drought stress triggers a significant enhancement of antioxidant enzyme activity. However, the application of the TiO₂NPs and AsA treatments, including 10 ppm TiO₂NPs, 50 ppm AsA, and 10 ppm TiO₂NPs combined with 50 ppm AsA, further increased SOD activity by 44%, 51%, 47%, and 27%, 48%, 18%, respectively (Figure 6).

CAT activity showed a significant increase when the coffee plants were treated with the various concentrations of TiO₂NPs and AsA, in comparison to those subjected to drought stress at 50% and 20% FC. Specifically, CAT activity rose by 36%, 70%, and 47% following the application of TiO₂NPs, while the AsA treatments led to increases of 21%, 66%, and 28%, respectively (Figure 6).

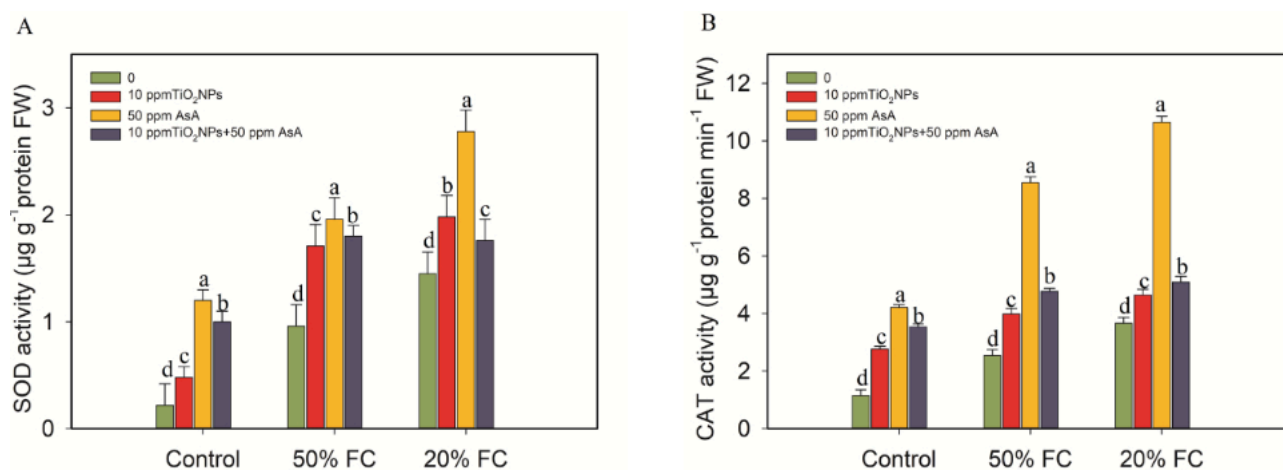


Figure 6 Impacts of TiO₂NPs and AsA on the SOD and CAT activity of coffee plants during drought stress conditions (50% FC and 20% FC). Statistically significant differences at the $p \leq 0.05$ level are indicated by bars with different letter annotations.

4. Discussion

Drought stress significantly restricts plant growth, resulting in a decrease in leaf water potential (ψ) (Yao et al., 2021; Waseem et al., 2021; Hasan et al., 2022; Liu et al., 2023). Conversely, when plants are treated with AsA and TiO₂NPs, their ability to withstand water scarcity is enhanced, mitigating the harmful impacts of this stress and preserving adequate turgor pressure in the leaves (Celi et al., 2023). The application of AsA and TiO₂NPs to coffee plants promotes increased drought tolerance in young coffee plants (Figure 7), as evidenced by the significant improvements in various growth parameters.

These results align with the findings reported by Ostadi et al. (2022) and Khazaei and Estaji (2020), both of which emphasize the important role that TiO₂NPs and AsA play in mitigating stress responses in plants (Figure 7). The RWC in leaves serves as an indicator of leaf hydration (Patanè et al., 2022). In our recent studies, as expected, the relative water content (RWC) reached its highest levels in control conditions and its lowest levels in conditions of drought. Nevertheless, TiO₂NPs and AsA resulted in an increase in RWC. Our analysis of the impact of TiO₂NPs and AsA on coffee plants in drought stress conditions revealed that the beneficial effects of these substances on

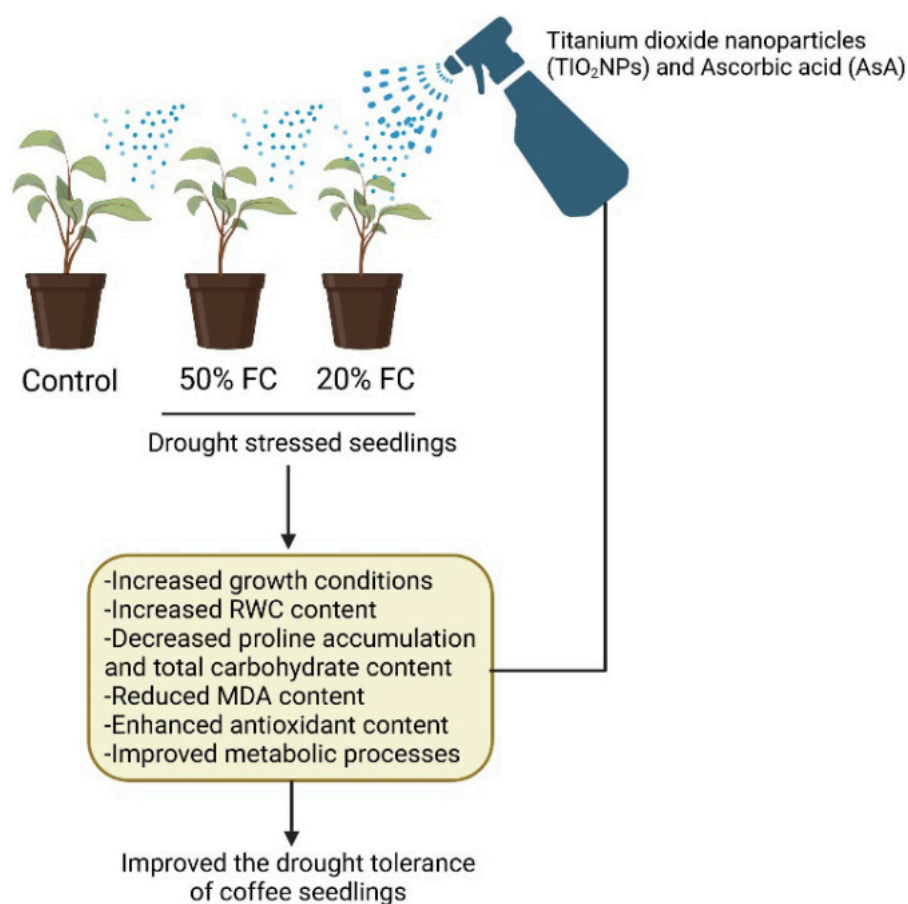


Figure 7 Schematic presentation of enhancement of the drought tolerance of coffee plants by titanium dioxide nanoparticles (TiO₂NPs) and ascorbic acid (AsA).

the studied characteristics can be attributed to their enhanced efficiency in water absorption. This, in turn, contributes to an improvement in turgor pressure and helps maintain water balance. The enhancement of RWC through the application of TiO₂NPs and AsA aligns with findings in studies involving sweet pepper (Khazaei et al., 2020) and sweet corn (Karvar et al., 2022), which share similarities with the current research.

Small organic molecules, like sugars, amino acids, organic and aromatic compounds, have the ability to control the cell's osmotic potential, which helps enhance water absorption during periods of drought stress (Jahan et al., 2023). Furthermore, these compounds serve to safeguard enzymes, biological membranes, and the photosynthetic system from oxidative harm (Khazaei et al., 2020). The current study demonstrates that coffee plants, when exposed to drought conditions, exhibit an increase in proline and soluble carbohydrates. In the context of drought-induced oxidative stress, there was an observed rise in proline levels in both pepper (Khazaei et al., 2020) and sweet corn (Karvar et al., 2022) plants and carbohydrate levels in pansy plants (Oraee et al., 2020), which is consistent with the findings in our present research. A related study demonstrated that multiple drought occurrences significantly affected the metabolic profile, including amino acids and carbohydrates, in coffee plants, indirectly aligning with our findings (Khazaei et al., 2020). Besides the amino acids and carbohydrates, other primary and secondary metabolites such as aromatic compounds and organic acids altered significantly in water-stress conditions, some previous studies were found parallel to our current results (Rodrigues et al., 2021; Guedes et al., 2018). In our study on coffee plants, the application of TiO₂NPs and AsA in drought conditions resulted in decreased levels of proline and carbohydrates. This reduction suggests that an alternative protective mechanism may be in operation,

potentially linked to the enhanced activation of antioxidative pathways by these treatments, which could mitigate oxidative stress more directly than osmotic adjustment.

In coffee plants, our study suggests that TiO₂NPs and AsA may influence the plant's oxidative balance by potentially modulating the enzymatic antioxidative defense system, rather than directly regulating already formed free radicals (Karamian et al., 2020). This could contribute to enhanced stress tolerance by mitigating the production of ROS, although direct measurements of ROS levels were not conducted in this study. The stability of the cell membrane is a crucial indicator of cellular integrity, especially in response to drought stress. The MDA content is a key marker for assessing cellular damage in plants, with elevated levels indicating increased membrane damage and compromised membrane integrity and function (Ahmed et al., 2023; Li et al., 2023; Sousa et al., 2023). In the present study, drought stress resulted in a notable rise in the MDA content, which indicates substantial disruption of the cell membrane. Nonetheless, the application of TiO₂NPs and AsA effectively mitigated these detrimental effects. The current findings indicate that TiO₂NPs and AsA play a critical role in safeguarding membrane stability in adverse environmental conditions. These observations are in line with previous studies on *Verbascum nudicuale* (Sousa et al., 2023) and pepper plants (Rodrigues et al., 2021), where TiO₂NPs and AsA were shown to enhance plant tolerance to drought-induced membrane damage.

A widely employed strategy for counteracting the damaging impacts of ROS in plants involves the utilization of antioxidant mechanisms. These mechanisms are designed to safeguard plants from oxidative stress and include enzymes like superoxide dismutase (SOD), which converts superoxide into hydrogen peroxide (H₂O₂), and other enzymes such as catalase and peroxidase that break down H₂O₂ into water (H₂O) and oxygen (O₂) (Abdelaal et al., 2022). Thus, the processes mitigating oxidative stress in plants are intricately linked to the action of SOD, which plays a pivotal role in activating other antioxidant enzymes (Hasan et al., 2021b). Our findings show a continuous increase in SOD enzyme activity in coffee plants, which corresponds with prior studies reporting similar increases under drought stress in coffee plants (Sousa et al., 2023) and soybeans (Hasan et al., 2020). CAT plays a crucial role in the glutathione-ascorbate cycle, which is vital for detoxifying H₂O₂ produced by the SOD enzyme in various cellular compartments. Our findings reveal that the activity of CAT in coffee plants exhibited similar patterns of change as those observed with SOD. When the coffee plants were treated with TiO₂NPs and AsA under drought stress, both SOD and CAT enzyme activities significantly increased. These results are consistent with earlier studies that showed that TiO₂NPs and AsA treatments enhanced antioxidant enzyme activity in corn (Karvar et al., 2022), cucumber (Zahra et al., 2021), and pepper (Khazaei et al., 2020).

Based on the experimental results, it is plausible that the application of TiO₂NPs and AsA led to an upregulation of both SOD and CAT activities, potentially linked to a reduction in the MDA content in the coffee leaves. This suggests that the combined use of TiO₂ nanoparticles (TiO₂NPs) and ascorbic acid (AsA) likely produces a synergistic effect by activating antioxidant defenses simultaneously. The enhanced activity of these key enzymes reflects their role in managing ROS and maintaining cellular stability, contributing to improved plant resilience in drought conditions. In addition, TiO₂NPs enhance water uptake by boosting root system efficiency, helping to maintain turgor pressure during drought. Meanwhile, AsA, as a potent antioxidant, plays a key role in scavenging reactive oxygen species (ROS), particularly by detoxifying hydrogen peroxide (H₂O₂) generated under drought-induced oxidative stress. This complementary interaction improving water regulation via TiO₂NPs and mitigating oxidative stress through AsA ultimately strengthens the drought tolerance of coffee plants.

5. Conclusion

In summary, our study has revealed that TiO₂NPs applied along with AsA leads to a substantial enhancement of physiological defense mechanisms in coffee plants. This combined application of TiO₂NPs and AsA effectively mitigates the negative impacts of water scarcity by increasing parameters like RWC, SOD, and CAT activities, thus

preventing oxidative damage. Given the limited existing literature on the joint use of TiO₂NPs and AsA via foliar application in coffee plants under drought stress and their protective role against oxidative damage, we propose that employing TiO₂NPs and AsA should be considered as a sustainable alternative strategy to enhance coffee production in drought conditions. Our findings could shed light on the beneficial impact of TiO₂NPs and AsA not only in coffee plants but also in other significant field crops. Deeper investigations into the molecular mechanisms by which TiO₂NPs and AsA improve drought tolerance in coffee plants are needed. They could involve gene expression analysis to comprehend how these treatments affect stress-related signaling pathways. Upcoming investigations should concentrate on the long-term effects of TiO₂NPs and AsA application on coffee as well as other agronomically important plants.

References

- Aebi, H. (1984). Catalase in vitro. In *Methods in Enzymology*; Elsevier; Vol. 105, pp. 121–126.
- Abdelaal, K., Alsubeie, M. S., Hafez, Y., Emeran, A., Moghanm, F., Okasha, S., Omara, R., Basahi, M. A., Darwish, D. B. E., & Ibrahim, M. F.M. (2022). Physiological and biochemical changes in vegetable and field crops under drought, salinity, and weeds stresses: Control strategies and management. *Agriculture*, 12, 2084. | <https://doi.org/10.3390/agriculture12122084>
- Ahmed, S. R., Anwar, Z., Shahbaz, U., Skalicky, M., Ijaz, A., Tariq, M. S., Zulfikar, U., Brestic, M., Alabdallah, N. M., & Alsubeie, M. S. (2023). Potential role of silicon in plants against biotic and abiotic stresses. *Silicon*, 15, 3283–3303. <https://doi.org/10.1007/s12633-022-02254-w>
- Alabdallah, N. M., Hasan, Md. M., Salih, A. M., Roushdy, S. S., Al-Shammari, A. S., Alsanie, S. I., & El-Zaidy, M. (2021). Silver nanoparticles improve growth and protect against oxidative damage in eggplant seedlings under drought stress. *Plant Soil and Environment*, 67, 617–624. <https://doi.org/10.17221/323/2021-PSE>
- Bakır, A. G., Bolat, I., Korkmaz, K., Hasan, Md. M., & Kaya, O. (2022). Exogenous nitric oxide and silicon applications alleviate water stress in apricots. *Life*, 12, 1454. <https://doi.org/10.3390/life12091454>
- Barrs, H. D. & Weatherley, P. E. (1962). A re-examination of the relative turgidity technique for estimating water deficits in leaves. *Australian journal of biological sciences*, 15(3), pp. 413–428.
- Bates, L. S. Waldren, R. P., & Teari, D. (1973). Rapid determination of free proline for water stress studies. *Plant Soil*, 39, 205–207.
- Castro, J. C., Castro, C. G., & Cobos, M. (2023). Genetic and biochemical strategies for regulation of L-ascorbic acid biosynthesis in plants through the L-galactose pathway. *Frontier in Plant Science*, 14, 1099829.
- Celi, G. E. A., Grato, P. L., Lanza, M. G. D. B., & Reis, A. R. D. (2023). Physiological and biochemical roles of ascorbic acid on mitigation of abiotic stresses in plants. *Plant Physiology and Biochemistry*, 202, 107970.
- Contreras-Oliva, A., Uscanga-Sosa, D. P., González-Rios, O., & Morales-Ramos, V. (2022). The use of coffee (*Coffea Arabica* L.) pulp in the preparation of a beverage with antioxidant properties. *International Food Research Journal*, 29, 274–282.
- DaMatta, F. M., Avila, R. T., Cardoso, A. A., Martins, S. C. V., & Ramalho, J. C. (2018). Physiological and agronomic performance of the coffee crop in the context of climate change and global warming: A review. *Journal of Agriculture and Food Chemistry*, 66, 5264–5274. <https://doi.org/10.1021/acs.jafc.7b04537>
- Dubois, M, Gilles, K. A., Hamilton, J. K., Rebers P. T., Smith, F. (1956). Colorimetric method for determination of sugars and related substances. *Analytical Chemistry*, 28(3), 350–6.
- Guedes, F. A. D. F., Nobres, P., Rodrigues Ferreira, D. C., Menezes-Silva, P. E., Ribeiro-Alves, M., Correa, R. L., DaMatta, F. M., & Alves-Ferreira, M. (2018). Transcriptional memory contributes to drought tolerance in coffee (*Coffea Canephora*) plants. *Environmental and Experimental Botany*, 147, 220–233. <https://doi.org/10.1016/j.envexpbot.2017.12.004>
- Hasan, Md. M., Alabdallah, N. M., Alharbi, B. M., Waseem, M., Yao, G., Liu, X.-D., Abd El-Gawad, H. G., El-Yazied, A. A., Ibrahim, M. F. M., & Jahan, M. S.(2021a). GABA: A key player in drought stress resistance in plants. *International Journal of Molecular Sciences*, 22, 10136. <https://doi.org/10.3390/ijms221810136>
- Hasan, Md. M., Alabdallah, N. M., Salih, A. M., Al-Shammari, A. S., ALZahrani, S. S., Al Lawati, A. H., Jahan, M. S., & Fang, X.-W. (2023). Modification of starch content and its management strategies in plants in response to drought and salinity: Current status

- and future prospects. *Journal of Soil Science and Plant Nutrition*, 23, 92–105.
<https://doi.org/10.1007/s42729-022-01057-7>
- Hasan, Md. M., Alharbi, B. M., Alhaithloul, H. A. S., Abdulmajeed, A. M., Alghanem, S. M., Al-Mushhin, A. A. M., Jahan, M. S., Corpas, F. J., Fang, X.-W., & Soliman, M. H. (2021b). Spermine-mediated tolerance to selenium toxicity in wheat (*Triticum Aestivum* L.) depends on endogenous nitric oxide synthesis. *Antioxidants*, 10, 1835.
<https://doi.org/10.3390/antiox10111835>
- Hasan, Md. M., Ali, Md. A., Soliman, M. H., Alqarawi, A. A., Abd_Allah, E. F., & Fang, X.-W. (2020). Insights into 28-homobrassinolide (HBR)-mediated redox homeostasis, AsA-GSH cycle, and methylglyoxal detoxification in soybean under drought-induced oxidative stress. *Journal of Plant Interactions*, 15, 371–385.
<https://doi.org/10.1080/17429145.2020.1832267>
- Hasan, Md. M., Liu, X.-D., Waseem, M., Guang-Qian, Y., Alabdallah, N. M., Jahan, M. S., & Fang, X.-W. (2022). ABA activated SnRK2 kinases: An emerging role in plant growth and physiology. *Plant Signaling & Behavior*, 17, 2071024.
<https://doi.org/10.1080/15592324.2022.2071024>
- Hasan, M. M., Alharby, H. F., Hajar, A. S., Hakeem, K. R., & Alzahrani, Y. (2018). Effects of magnetized water on phenolic compounds, lipid peroxidation and antioxidant activity of *Moringa* species under drought stress. *Journal of Animal & Plant Sciences*, 28.
- Heath, R. L., & Packer, L. (1968). Photoperoxidation in isolated chloroplasts. *Archives of Biochemistry and Biophysics*, 125, 189–198.
[https://doi.org/10.1016/0003-9861\(68\)90654-1](https://doi.org/10.1016/0003-9861(68)90654-1)
- Jahan, M. S., Guo, S., Sun, J., Shu, S., Wang, Y., El-Yazied, A. A., Alabdallah, N. M., Hikal, M., Mohamed, M. H. M., Ibrahim, M. F. M., & Hasan M. M. (2021). Melatonin-mediated photosynthetic performance of tomato seedlings under high-temperature stress. *Plant Physiology and Biochemistry*, 167, 309–320.
<https://doi.org/10.1016/j.plaphy.2021.08.002>
- Jahan, M. S., Hasan, M. M., Alotaibi, F. S., Alabdallah, N. M., Alharbi, B. M., Ramadan, K. M. A., Bendary, E. S. A., Alshehri, D., Jabborova, D., & Al-Balawi, D. A. (2022). Exogenous putrescine increases heat tolerance in tomato seedlings by regulating chlorophyll metabolism and enhancing antioxidant defense efficiency. *Plants*, 11, 1038.
<https://doi.org/10.3390/plants11081038>
- Jahan, M. S., Li, G., Xie, D., Farag, R., Hasan, Md. M., Alabdallah, N. M., Al-Harbi, N. A., Al-Qahtani, S. M., Zeeshan, M., & Nasar, J. et al. (2023). Melatonin mitigates salt-induced growth inhibition through the regulation of carbohydrate and nitrogen metabolism in tomato seedlings. *Journal of Soil Science and Plant Nutrition*, 23, 4290–4308.
<https://doi.org/10.1007/s42729-023-01348-7>
- Jovanovc, B. (2015). Critical review of public health regulations of titanium dioxide, a human food additive. *Integrated Environmental Assessment and Management*, 11, 10–20.
- Karvar, M., Azari, A., Rahimi, A., Maddah-Hosseini, S., & Ahmadi-Lahijani, M. J. (2022). Titanium dioxide nanoparticles (TiO₂-NPs) enhance drought tolerance and grain yield of sweet corn (*Zea Mays* L.) under deficit irrigation regimes. *Acta Physiologiae Plantarum*, 44, 14. <https://doi.org/10.1007/s11738-021-03349-4>
- Karamian, R., Ghasemlou, F., & Amiri, H. (2020). Physiological evaluation of drought stress tolerance and recovery in *Verbascum Sinuatum* plants treated with methyl jasmonate, salicylic acid, and titanium dioxide nanoparticles. *Plant Biosystems – An International Journal Dealing with all Aspects of Plant Biology*, 154, 277–287.
<https://doi.org/10.1080/11263504.2019.1591535>
- Khazaei, Z., Esmaelpour, B., & Estaji, A. (2020). Ameliorative effects of ascorbic acid on tolerance to drought stress in pepper (*Capsicum Annuum* L.) plants. *Physiology and Molecular Biology of Plants*, 26, 1649–1662. <https://doi.org/10.1007/s12298-020-00846-7>
- Khazaei, Z., & Estaji, A. (2020). Effect of foliar application of ascorbic acid on sweet pepper (*Capsicum Annuum*) plants under drought stress. *Acta Physiologiae Plantarum*, 42, 118. <https://doi.org/10.1007/s11738-020-03106-z>
- Khan, A., Anwar, Y., Hasan, M., Iqbal, A., Ali, M., Alharby, H. F., Hakeem, K. R., & Hasanuzzaman, M. (2017). Attenuation of drought stress in Brassica seedlings with exogenous application of Ca²⁺ and H₂O₂. *Plants*, 6, 20.
- Li, J., Arkorful, E., Cheng, S., Zhou, Q., Li, H., Chen, X., Sun, K., & Li, X. (2018). Alleviation of cold damage by exogenous application of melatonin in vegetatively propagated tea plant (*Camellia sinensis* (L.) O. Kuntze). *Scientia Horticulturae*, 238, 356–362.
<https://doi.org/10.1016/j.scienta.2018.04.068>
- Li, X., Ghanizadeh, H., Han, Z., Wang, Q., Li, F., Qiu, Y., Zhang, Y., Chen, X., Liu, J., & Wang, A. (2023). Metabolic profile and physiological mechanisms underlying the promoting effects of TiO₂ NPs on the photosynthesis and growth of tomato. *Scientia Horticulturae*, 322, 112394. <https://doi.org/10.1016/j.scienta.2023.112394>

- Liu, X., Zeng, Y., Zhang, X., Tian, X., Hasan, Md. M., Yao, G., & Fang, X. (2023). Polyamines inhibit abscisic acid-induced stomatal closure by scavenging hydrogen peroxide. *Physiologia Plantarum*, 175, e13903. <https://doi.org/10.1111/ppl.13903>
- Melo, T. M., Schauerte, M., Bluhm, A., Slány, M., Paller, M., Bolan, N., Bosch, J., Fritzsche, A., & Rinklebe, J. (2022). Ecotoxicological effects of per- and polyfluoroalkyl substances (PFAS) and of a new PFAS adsorbing organoclay to immobilize PFAS in soils on earthworms and plants. *Journal of Hazardous Materials*, 433, 128771.
- Menard, A., Drobne, D., & Jemec, A. (2011). Ecotoxicity of nanosized TiO₂: Review of in vivo data. *Environmental Pollution*, 159, 677–684.
- Merga, D., & Beksisa, L. (2023). Mechanisms of drought tolerance in coffee (*Coffea Arabica* L.): Implication for genetic improvement program. *American Journal of BioScience*, 11, 63–70.
- Menezes-Silva, P. E., Sanglard, L. M. V. P., Ávila, R. T., Morais, L. E., Martins, S. C. V., Nobres, P., Patreze, C. M., Ferreira, M. A., Araújo, W. L., & Fernie, A. R. (2017). Photosynthetic and metabolic acclimation to repeated drought events play key roles in drought tolerance in coffee. *Journal of Experimental Botany*, 68, 4309–4322. <https://doi.org/10.1093/jxb/erx211>
- Oraee, A., & Tehranifar, A. (2020). Evaluating the potential drought tolerance of pansy through its physiological and biochemical responses to drought and recovery periods. *Scientia Horticulturae*, 265, 109225. <https://doi.org/10.1016/j.scienta.2020.109225>
- Ostadi, A., Javanmard, A., Amani Machiani, M., Sadeghpour, A., Maggi, F., Nouraein, M., Morshedloo, M. R., Hano, C., & Lorenzo, J. M. (2022). Co-application of TiO₂ nanoparticles and arbuscular mycorrhizal fungi improves essential oil quantity and quality of sage (*Salvia Officinalis* L.) in drought stress conditions. *Plants*, 11, 1659. <https://doi.org/10.3390/plants11131659>
- Patanè, C., Cosentino, S. L., Romano, D., & Toscano, S. (2022). Relative water content, proline, and antioxidant enzymes in leaves of long shelf-life tomatoes under drought stress and rewatering. *Plants*, 11, 3045. <https://doi.org/10.3390/plants11223045>
- Rahman, M. A., Woo, J. H., Song, Y., Lee, S.-H., Hasan, M. M., Azad, M. A. K., & Lee, K.-W. (2022). Heat shock proteins and antioxidant genes involved in heat combined with drought stress responses in perennial rye grass. *Life*, 12, 1426. <https://doi.org/10.3390/life12091426>
- Rodrigues, A. M., Jorge, T., Osorio, S., Pott, D. M., Lidon, F. C., DaMatta, F. M., Marques, I., Ribeiro-Barros, A. I., Ramalho, J. C., & António, C. (2021). Primary metabolite profile changes in *Coffea* spp. promoted by single and combined exposure to drought and elevated CO₂ concentration. *Metabolites*, 11, 427. <https://doi.org/10.3390/metabo11070427>
- Sheikhalipour, M., Gohari, G., Esmaielpour, B., Panahirad, S., Milani, M. H., Kulak, M., & Janda, T. (2023). Melatonin and TiO₂ NPs application-induced changes in growth, photosynthesis, antioxidant enzymes activities, and secondary metabolites in *Stevia* (*Stevia Rebaudiana* Bertoni) under drought stress conditions. *Journal of Plant Growth Regulation*, 42, 2023–2040. <https://doi.org/10.1007/s00344-022-10679-1>
- Silva, P. C. D., Junior, W. Q. R., Ramos, M. L. G., Rocha, O. C., Veiga, A. D., Silva, N. H., Brasileiro, L. D. O., Santana, C. C., Soares, G. F., & Malaquias, J. V. (2022). Physiological changes of Arabica coffee under different intensities and durations of water stress in the Brazilian Cerrado. *Plants*, 11, 2198. <https://doi.org/10.3390/plants11172198>
- Sousa, G. F. D., Silva, M. A., Carvalho, M. R. D., Morais, E. G. D., Benevenuto, P. A. N., Van Opbergen, G. A. Z., Van Opbergen, G. G. A. Z., & Guilherme, L. R. G. (2023). Foliar selenium application to reduce the induced-drought stress effects in coffee seedlings: Induced priming or alleviation effect? *Plants*, 12, 3026. <https://doi.org/10.3390/plants12173026>
- Wang, J., Wang, Z., Wang, W., Wang, Y., Hu, X., Liu, J., Gong, X., Miao, W., Ding, L., & Li, X. (2022). Synthesis, modification and application of titanium dioxide nanoparticles: A review. *Nanoscale*, 14, 6709–6734.
- Waseem, M., Nie, N. F., Yao, G. Q., Hasan, M. M., Xiang, Y., & Fang, X. W. (2021). Dew absorption by leaf trichomes in *Caragana korshinskii*: An alternative water acquisition strategy to withstand drought in an arid environment. *Physiologia Plantarum*, 172, 528–539.
- Yao, G., Nie, Z., Zeng, Y., Waseem, M., Hasan, Md. M., Tian, X., Liao, Z., Siddique, K. H. M., & Fang, X. (2021). A clear trade-off between leaf hydraulic efficiency and safety in an aridland shrub during regrowth. *Plant, Cell & Environment*, 44, 3347–3357. <https://doi.org/10.1111/pce.14156>
- Zahra, G., Marjan, M., Taher, B., & Ranjbar, M. E. (2021). Foliar application of ascorbic acid and gamma aminobutyric acid can improve important properties of deficit irrigated cucumber plants (*Cucumis Sativus* Cv. Us). *Gesunde Pflanzen*, 73, 77–84.