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## ORIGINAL RESEARCH

# Chemical composition and anti-inflammatory and anticancer effects of methanolic extract of *Arum palaestinum* spadices

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## Abstract

*Arum palaestinum* is a wild edible plant that belongs to the Araceae family and grows in different regions of the Middle East. The present study aimed to investigate the chemical composition and bioactivities of *A. palaestinum* spadices utilizing GC/MS analysis of the non-polar fraction, the MTS assay as well as membrane stabilization and disk diffusion tests on various methanolic extracts. Twelve known compounds from the non-polar fraction were characterized by the GC/MS analysis. The spathe extract exhibited the maximum inhibition activity against HeLa cells and good activity against HepG-2 cells. The spathe extract showed a relatively high selectivity ratio towards three cancer cell lines, i.e., around 0.22, 0.73, and 0.16 towards HeLa, MCF-7, and HepG-2, respectively. Membrane stabilization was induced by all the crude extracts, ranging between a 70.7% hemolysis inhibition rate exerted by the spathe extract and 75.3% inhibition by the XAD extract. The results suggest that *A. palaestinum* spadices may be considered as a potential source of new bioactive agents for drug development.

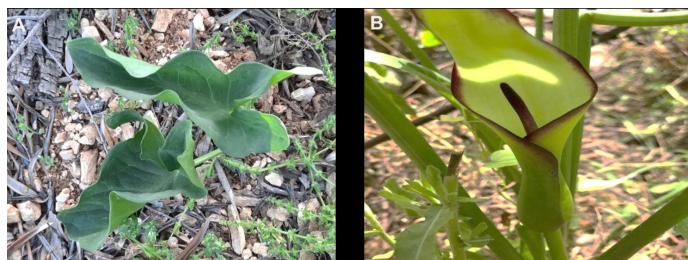
## Keywords

*Arum palaestinum*; chemical composition; methanolic extracts; anticancer; anti-inflammatory

## 1. Introduction

The screening of flora for biological active compounds started in the late nineties (Husein et al., 2014b) and is still being considered nowadays. *A. palaestinum* Boiss is one of the 26 species of flowering plants of the *Arum* genus that belongs to the Araceae family (Jaber et al., 2020), which has the highest diversity in the Mediterranean region (Farid et al., 2015). It is widely used as a traditional folk medicinal plant in Arabic and Greek traditions (Máthé & Bandoni, 2021). It has been utilized for treatment of several diseases, such as breast, liver, and colon cancer (Jaradat et al., 2016).

The interest in the *A. palaestinum* plant is increasing for its bioactive phytochemicals and secondary metabolites that have been discovered and are still being under discovery (El-Desouky et al., 2014; Farid et al., 2015). The leaf extract containing various phytochemical metabolites: flavonoids, phenolic acids, and alkaloids (Abu-Reidah et al., 2015) was reported to have antidiabetic (Hawash et al., 2020) anti-inflammatory (Abu-Qatouseh et al., 2019), and analgesic activities (Qnais et al., 2017) as well as anti-cancer and antioxidant activities against different cancer cell lines (Jaradat & Abualhasan, 2016). Its extracts were reported to have anti-tumour cytotoxicity towards



**Figure 1** *A. palaestinum* leaves (A) and spadix surrounded by spathes (B).

different cancer cells (Aboul-Enein et al., 2012; Hatmal et al., 2017). Furthermore, its synergistic combinations have been reported to have a promising therapeutic capability against cancer (Booth et al., 2021).

Based on these considerations, we came to the realization that the bioactivity of the flowering parts of *A. palaestinum* (Figure 1) is yet to be reported. Therefore, we assumed that the examination of bioactivities of *A. palaestinum* flowering parts would be extremely beneficial and would have a potential application in the treatment of several diseases. In this work, we aimed to study the chemical composition of the non-polar fraction of *A. palaestinum* spadices and the bioactivities of different extracts of the flowering part. The methanolic extracts of spadices and spathes in addition to the aqueous extract of spadices were evaluated for their anti-inflammatory and anticancer activity against MCF-7, HeLa, Hep-G2, and LX-2 cell lines.

## 2. Material and methods

### 2.1. Chemicals and equipment

Methanol 99% (MeOH), trifluoroacetic acid (TFA), DEME media, fetal bovine serum (99.5%), penicillin-streptomycin (99%), phosphate buffer saline (PBS), trypsin (99.5%), DMSO (99%), Doxorubicin (99.9%), and 5-Fluorouracil (99.9%) were all purchased from Sigma-Aldrich.

### 2.2. Collection of plant material

*A. palaestinum* (Voucher number BERC-C0064) flowers were collected from Ramallah, West Bank/Palestine during March/April 2021, air dried in a dark place, and preserved at the chemistry labs in Birzeit University, Ramallah/Palestine. The dark purple spadices and spathes were separated, ground, and weighed.

### 2.3. Extraction of plant material

The samples of dried *A. palaestinum* spadices (41.4880 g) and *A. palaestinum* spathes (35.3189 g) were extracted three times over night at 4 °C with 400 mL of 10% distilled water H<sub>2</sub>O in MeOH (v/v; containing 0.5% trifluoroacetic acid, TFA). The combined solutions were filtrated, concentrated under reduced pressure in order to remove the solvent at 35 °C, weighed, and dried completely with Nitrogen to yield 23.3274 g and 14.9934 g respectively (Husein et al., 2014a).

The nonpolar compounds were removed from the *A. palaestinum* extract by partitioning with 120 mL DW against hexane. The aqueous phase was subjected to Amberlite XAD-7 column chromatography (Hamann, 2006) using 10% distilled water H<sub>2</sub>O in MeOH (v/v; containing 0.5% TFA) as an eluent to yield 2.0751 g. A sample of dried XAD-7 purified extract was taken for anticancer studies.

A 109.7852 g powder sample of the flowering parts of the plant was soaked twice, each in 620 mL DCM (Devi et al., 2020), basified with ammonium hydroxide (NH<sub>4</sub>OH) until the solution pH 8–9, filtrated, and concentrated to yield 2.0096 g of crude extract. Another 48.9208 g of the mixture of powdered flowering parts was soaked twice for four hours, each in 300 mL ethyl acetate, basified with NH<sub>4</sub>OH (until pH 9), concentrated, and dried to yield 0.7352 g of concentrated extract.

## 2.4. GC/MS analysis

The GC/MS analysis (Kaur & Arora, 2015; Zargoun et al., 2020) of volatile compounds was performed on a *PerkinElmer Clarus 600* mass spectrometer using a non-polar 5% phenyl methylpolysiloxane TG-5MS capillary column (30.00 m × 0.25 mm × 0.25 mm) in the following conditions: oven temperature program from 80 °C (3 min) to 280 °C at 10 C/min, then isothermal treatment at 280 °C for 3 min. The carrier gas was helium at a flow rate of 1 mL/min. The volume of injected sample was 10 µL of the sample in acetonitrile using a split injection technique (1:10), ionization energy 70 eV, in the electronic ionization mode.

## 2.5. *In-vitro* cytotoxicity MTS assay and anticancer activity

### 2.5.1. Cell culture

Four ATCC adenocarcinoma cell lines were used in this study: MCF-7 (human breast cancer cell line), HeLa (immortalized cervical cancer cells), HepG-2 (hepatocellular carcinoma), and LX-2 (normal human hepatic stellate cells).

The cell lines were grown in RPMI-1640 medium supplemented with antibiotics, antimycotics, L-glutamine, and fetal bovine serum. The cells were incubated at 37 °C in a humid chamber containing 5% CO<sub>2</sub>. A total of  $2.5 \times 10^4$  cells were seeded in each well of flat 96-well plates and incubated for 72 hours until almost confluent. The cells were then washed with Hank's balanced salt solution and fresh medium was added. The cells were then treated with different concentrations (250, 500, 2,000 and 4,000 µg/mL) of the three *A. palaestinum* extracts: Spathe, Spadix, and XAD extracts and incubated as mentioned above.

### 2.5.2. Cell viability assay

After incubation, the viability of the cells was investigated by the *Cell Titer 96® Aqueous One Solution Cell Proliferation* (MTS) assay according to the manufacturer's instructions (Promega Corporation, Madison, WI) (Shi et al., 2009). 20 µL of MTS solution per 100 µL of media were added to each well and kept at 37 °C for 2–4 hours. Then, the absorbance was measured at 490 nm with an ELISA reader. The IC<sub>50</sub> values of the extracts were determined by plotting the cell viability percentage versus the working concentrations of each extract of the four cell lines separately.

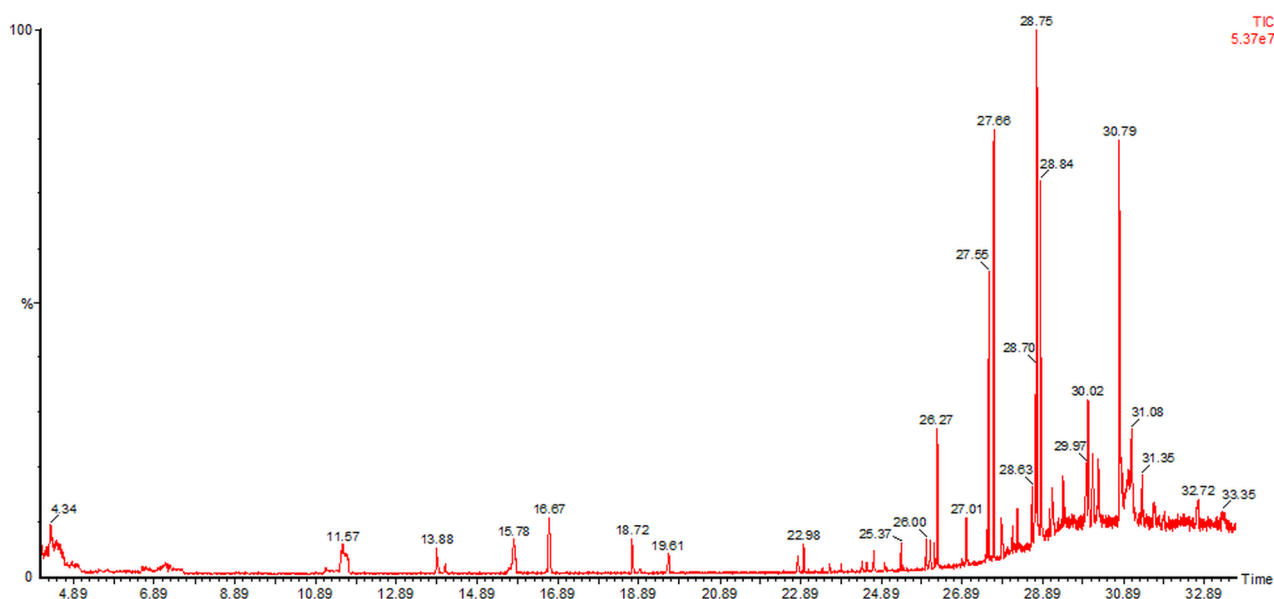
### 2.5.3. *In-vitro* anti-inflammatory assay

Blood samples (5 ml) from healthy human volunteers who had not taken any non-steroidal anti-inflammatory drugs (NSAIDs) for 2 weeks were collected. After washing with 0.9% normal saline and centrifuged for 10 min at 2,000 rpm, the volume of the blood was reconstituted as a 40% (v/v) suspension with an isotonic buffer solution (10 mM sodium phosphate buffer pH 7.4, NaH<sub>2</sub>PO<sub>4</sub> (0.2 g/l), Na<sub>2</sub>HPO<sub>4</sub> (1.15 g/l), and NaCl (9.0 g/l)). Standard solutions of acetylsalicylic acid (25–0.195 mg/mL) were prepared by dissolving 100 mg of acetylsalicylic in 2 mL of a hypotonic solution. Blood samples containing the hypotonic solution (154 mM NaCl in 10 mM Sodium Phosphate Buffer at pH 7.4) and the extracts of *A. palaestinum* spadices, spathes, and XAD were prepared in the same range of acetylsalicylic acid concentrations. Washed red blood cells 10 µm (0.01 mL) were added to each concentration of the control, i.e., the standard hypotonic solution with RBC suspension. The experiment was carried out twice. The mixture was incubated for 10 min at room temperature and centrifuged at 3,000 rpm. The absorbance of the supernatant was measured at 540 nm.

## 3. Results

### 3.1. GC/MS analysis

The DCM and EtOAc fractions of *A. palaestinum* were subjected to GC/MS analysis to identify volatile compounds (Figure 2, Figure S1). A total of twelve major compounds were identified (Table 1).



**Figure 2** GC/MS chromatogram of the non-polar fraction of *A. palaestinum* spadices (DCM extract).

**Table 1** GC-MS analysis of DCM and EtOAc fractions of the flowering part of *A. palaestinum*.

Extract	Compounds	Molar mass (g/mol)	Retention time (min)	Molecular formula
DCM	Octadecanamide	283.5	28.75	C <sub>18</sub> H <sub>37</sub> NO
	9-Z-Octadecenamide	281.48	30.79	C <sub>18</sub> H <sub>35</sub> NO
	Squalane	244.8	27.55	C <sub>30</sub> H <sub>62</sub>
	Hexadecanamide	255.44	28.84	C <sub>16</sub> H <sub>33</sub> NO
	Octadecanal	268.49	26.27	C <sub>18</sub> H <sub>36</sub> O
	(Z)-13-Docosenamide	337.58	30.79	C <sub>22</sub> H <sub>43</sub> NO
	Heptacosane	380.7	27.66	C <sub>27</sub> H <sub>56</sub>
	Isocitronellol	156.27	19.61	C <sub>10</sub> H <sub>20</sub> O
	3,5-bis(1,1-dimethylethyl)-Phenol	206.32	16.67	C <sub>14</sub> H <sub>22</sub> O
	EtOAc	Hexadecanoic acid methyl ester	270.45	24.67
(E,E)-9,12-Octadecadienoic acid methyl ester		294.5	26.21	C <sub>19</sub> H <sub>34</sub> O <sub>2</sub>
Linoleic acid ethyl ester		308.5	26.72	C <sub>20</sub> H <sub>36</sub> O <sub>2</sub>

### 3.2. *In-vitro* bioactivity assays of *A. palaestinum* extracts

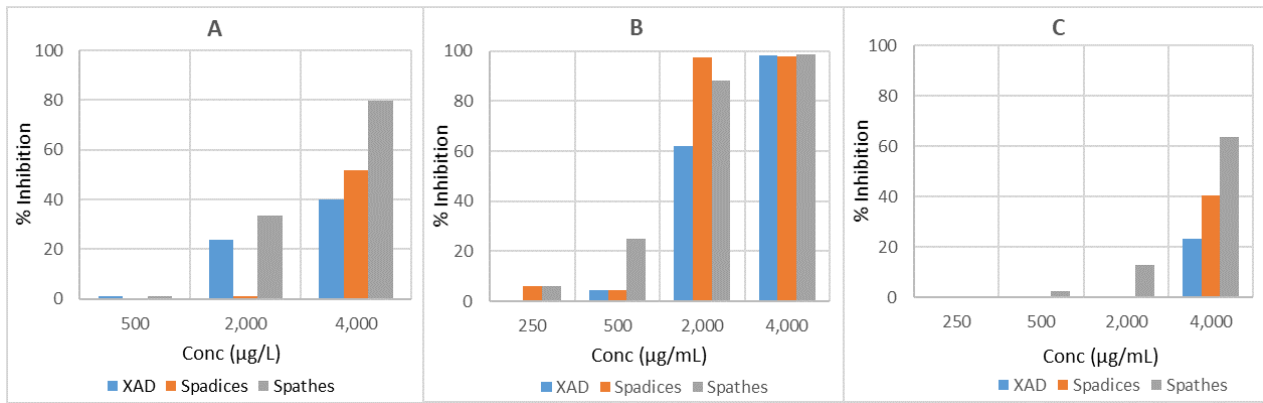
#### 3.2.1. Anticancer activity of *A. palaestinum* extracts

The MTS assay was done on HepG2, HeLa, and MCF7 cells as well as normal cells LX-2 to assess the anticancer effects of *A. palaestinum* spathe, spadix, and XAD extracts using four concentrations (250, 500, 2,000, and 4,000 µg/mL) (Figure 3, Table S1, Table S2, Table S3). Their IC<sub>50</sub> values were evaluated (Table 2) in addition to their selectivity ratios (Figure 4).

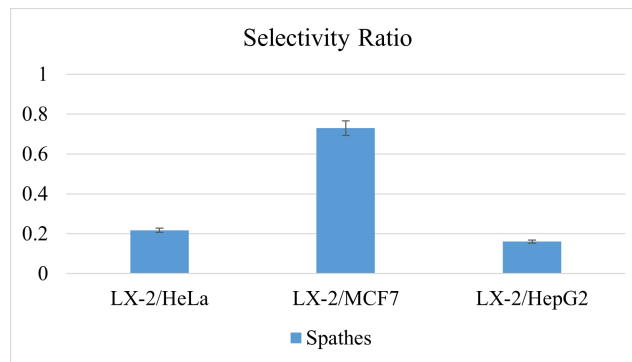
#### 3.2.2. Anti-inflammatory activity of *A. palaestinum* extracts

The anti-inflammatory activity was studied using human blood samples collected from healthy volunteers who had not used any NSAIDs for 2 weeks, and it was done as established by Shahriar et al. (2015). The percentage of RBC membrane stabilization or inhibition of hemolysis was calculated by the following equation (Miah et al., 2018) (Figure 5, Table S4, Table S5, Table S6, Table S7).

$$\% \text{ Inhibition of hemolysis} = 100 \cdot [\text{OD}_1 - \text{OD}_2 / \text{OD}_1].$$



**Figure 3** The percent inhibition of HeLa (A), MCF7 (B), and HepG2 (C) cancer cell lines using different concentrations of *A. palaestinum* extracts.



**Figure 4** Selectivity ratio for the Spathes extract ( $IC_{50}$  of LX-2 to  $IC_{50}$  of HeLa and MCF7 and HepG2 cancer cell lines).

**Table 2**  $IC_{50}$  of *A. palaestinum* extracts against cancer cell lines.

	$IC_{50}$ (µg/mL)		
	HeLa	MCF7	HepG2
XAD	NI	1,893.51±3.78	NI
Spadices	3,911.99±3.70	818.24±2.14	NI
Spathes	2,530.04±2.47	751.17±3.01	3,412.25±3.66
Doxorubicin	1.598±0.67	1.21±1.0	0.84±1.1
5-Fluorouracil	0.036±0.01	0.056±0.44	0.063±0.50

OD<sub>1</sub> – optical density of hypotonic buffered saline solution alone.

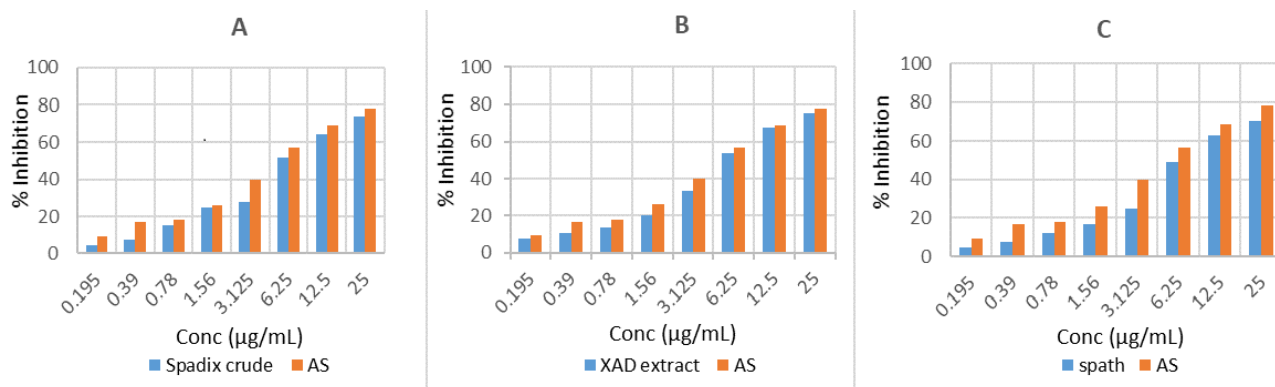
OD<sub>2</sub> – optical density of test sample in hypotonic solution.

#### 4. Discussion

The taxonomic identification of the plant material was confirmed by Prof. Munir Naser from Birzeit University. The voucher specimen (*Arum palaestinum* Boiss. Voucher No. BERC-BZ-001) has been deposited at BERC Herbarium, Til, Nablus, Palestine.

##### 4.1. GC/MS analysis

Twelve known compounds in the non-polar fraction of *A. palaestinum* were identified by the GC/MS analysis, which have been recorded to have recognized biological activ-



**Figure 5** The percent inhibition of hemolysis (A) *A. palaestinum* spadice crude extract (B) XAD fraction of *A. palaestinum* spadices (C) *A. palaestinum* spathe crude extract.

ity. For example, *Heptacosane* was reported to modulate the activity of P-glycoprotein and exert synergistic effects side by side to the Doxorubicin drug, enhancing its cytotoxic activity against HL-60R cells (Labbozzetta et al., 2022). In addition, *3,5-bis(1,1-dimethylethyl)-Phenol* is considered an anticancer agent that is still under study (Rizvi et al., 2014), whereas *(Z)-13-Docosenamamide* was reported to exhibit anti-fungal and anti-tumor properties (Dai et al., 2020). In turn, hexadecanoic acid, a well-known saturated fatty acid, has recently attracted attention and is recently marketed as a health supplement; its methyl ester derivatives are assigned much biological importance, as they have antioxidant, anticancer, antihistaminic, and anti-malarial properties (Melariri et al., 2012). Furthermore, *Hexadecanoic acid methyl ester* has recently been reported to have antibacterial effects against pathogenic bacteria, such as *P. aeruginosa*, *Klebsiella pneumoniae*, and *Bacillus subtilis* (Shaaban et al., 2021). Squalenes have gained interest as components in cosmetic pharmaceuticals, and they are being under recent *ex-vivo* studies to evaluate their ability to enhance the physiological functions of different polyphenols in order to increase their skin permeability and improve their antiaging effects (Oliveira et al., 2022).

#### 4.2. *In-vitro* bioactivity assays of *A. palaestinum* extracts

##### 4.2.1. Anticancer activity of *A. palaestinum* extracts

Concentrations in the range of 250–4,000 µg/mL of the extracts were used to determine the optimum concentration of the extract exhibiting significant anticancer activity against the cancer cell lines investigated in this study. The specific concentrations are therefore determined through empirical testing and may vary depending on the goals and methodology of the extraction process. Common concentrations used in anticancer studies of plant extracts can be up to 500 µg/mL (Halecki et al., 2022; Ho et al., 2022; Nisa et al., 2022). Testing various concentrations is a standard strategy to determine the optimal concentration of a plant extract, where maximum anticancer activity is achieved while minimizing potential harm to normal cells. Furthermore, initial investigations were carried out to evaluate the cytotoxic impact of plant extracts on normal cells prior to proceeding to subsequent experiments. This precautionary step ensures that the plant extract selectively affects cancer cells without causing substantial harm to healthy cells. In this study, normal cells LX-2 (LX-2 human hepatic stellate cell line) were used and it was found that the extracts used in this study exerted weak or negligible anticancer effects on these human normal skin cells.

Based on the above results, it can be claimed that the spathe extract has the maximum inhibition activity against HeLa cells, which reached about 80% inhibition at 4,000 µg/mL, while the other extracts showed lower inhibition. The extracts of XAD, spadices, and spathes showed significant anticancer activity (more than 98% inhibition) against the MCF-7 cancer cell line at 4,000 µg/mL. These extracts showed moderate to strong inhibitory activity towards the same cancer cells at 2,000 µg/mL, while, low inhibition activity was observed at the concentration of 500 µg/mL.



The 4,000 µg/mL concentration showed anticancer activity of all extracts against HepG-2 cancer cells. 63.7% inhibition was exerted by the spathe extract, while 40.5% and 23.3% inhibition was induced by the spadice and XAD extracts, respectively.

All the extracts had higher IC<sub>50</sub> values compared to the positive controls Doxorubicin and 5-Fluorouracil against the HeLa and MCF-7 cancer cell lines, which indicates a lower inhibition ability of the *A. palaestinum* plant extracts compared to the aforementioned drugs. However, only the spathe extract had good activity against the HepG-2 cells in comparison with the other extracts. Noteworthy, the *A. palaestinum* extracts displayed anticancer properties against the three cancer cell lines. Moreover, these extracts were shown to have weak or negligible anticancer effects on human normal skin fibroblast cells LX-2, which is promising evidence in the field of cancer treatment by plant extracts. Nonetheless, their cytotoxic concentrations are high and controversial. Since several extracts from the aerial parts of this plant have long been claimed to exhibit anticancer and antiproliferative action with very low IC<sub>50</sub> values, more research studies are required to validate and strengthen these findings.

#### 4.2.2. Selectivity ratio of the extracts

Based on the fact that the spathe extract yielded the most encouraging results, its effects on normal cells was investigated. Therefore, the selectivity ratio, i.e., % inhibition of the extract of LX-2 to the HeLa, MCF-7, and HepG-2 cancer cell lines, was calculated. The selectivity ratio of the extract was around 0.22 for LX-2 to HeLa, 0.73 for LX-2 to MCF7, and 0.16 for LX-2 to HepG2. If these ratios were compared to those of the positive controls (Dox and 5-FU, which have lower selectivity ratios), the spathe extract could be considered to have possible crucial action against the three aforementioned cancer cell lines. It is important to notice that as the selectivity ratio of a compound or extract increases, it is considered safe compared to compounds with low selectivity factors like Dox and 5-FU.

In this regard, it is widely recognized that certain portions of this plant have anticancer properties. Its ethanolic and ethyl acetate extracts, for example, have been shown to inhibit the growth of MFC-7 cancer cells (El-Desouky et al., 2007; Husein et al., 2014a). Furthermore, a number of its isolated constituents and distinct extracts have been found to exhibit detectable actions against a variety of cancer cell lines, including the cell lines examined in this work: cervix (HeLa), liver (HepG2), and breast (MFC7) carcinoma cell lines (Farid et al., 2015, 2017; Hatmal et al., 2017; Naseef et al., 2017). Similarly, the encouraging findings of the current study, which focuses on the flowering parts of the plant, enhance the bioactive potential of *A. palaestinum* as a whole medicinal plant.

#### 4.2.3. Anti-inflammatory activity of *A. palaestinum* extracts

All the *A. palaestinum* extracts function similarly towards RBC membrane stabilization. They exerted strong activity against lysis with IC<sub>50</sub> of 6.25 mg/mL and prevented the hemolysis of the blood cells (induced by the hypotonic solution) up to 75%, compared to acetyl salicylic acid (AS) (78% inhibition at 25 mg/mL). The maximum anti-inflammatory effect was significant at the 25 mg/mL concentration of all the *A. palaestinum* extracts, which induced more than 70% inhibition. Higher inhibition was achieved by the XAD extract of *A. palaestinum*. Aqueous extracts could be attributed with increased flavonoid contents (Farid et al., 2015). The anti-inflammatory results of the flowering parts are contrary to the findings for leaf extracts documented by Khalaf et al. (2015), who reported that arum leaves extracts did not exhibit any considerable anti-inflammatory activity. On the other hand, it exerted a unique anti-inflammatory effect, as reported in previous work (Abu-Qatouseh et al., 2019). To the best of our knowledge, this is the first study to investigate the bioactivity of *A. palaestinum* flowering parts, as all of the previous studies were focused on its aerial parts, especially leaves. In this regard, it should be pointed out that these findings are in agreement with literature reports on the anticancer activity of this plant. Some of its alcoholic and organic, as well as aqueous, extracts have been reported to have anticancer activity against several studied cancer cell lines, including cancer cells that were studied in this work (Azab, 2023).

Extracts from *A. palaestinum* leaves have more potent anti-cancer effects, compared to plant's flowering parts, as indicated by the IC<sub>50</sub> values found in prior literature. This underscores the necessity for additional research on plant's spadices and spathes. Moreover, future studies on combinatorial extracts of plant's aerial and flowering parts are recommended to minimize their active concentrations. It is noteworthy to point out that all extracts were examined for their antibacterial and antifungal activity. No extracts showed any growth inhibition when applied to both Gram-negative bacteria (*Escherichia coli* and *Pseudomonas aeruginosa*) and Gram-positive bacteria (*Staphylococcus aureus*) at all concentrations ranging from 0.78 mg/mL to 100 mg/mL. In the same manner, they exhibited no inhibition activity on the fungi *Candida albicans* at the same aforementioned concentrations.

## 5. Conclusion

The present study is considered to be the first report on anticancer cytotoxic activities exhibited by extracts from *A. palaestinum* spadices to provide evidence for its anti-tumor activities. The cytotoxicity assay of different extracts on the tested cancer cell lines (HeLa, MCF-7, and HepG-2) showed variations in their effects. The examined extracts exhibited concentration-dependent inhibitory activity against the examined cancer cell lines but weak discernible cytotoxic effects. Despite the high cytotoxic concentration, the spathe extracts showed the best inhibitory effects and a relatively high selectivity ratio, which reveals possible crucial action against the cancer cell lines. Nevertheless, the present study showed strong and comparable anti-inflammatory effects of *A. palaestinum* spadice extracts. Since the GC/MS analysis suggested the potential presence of many bioactive compounds with varying degrees of bioactivity, more investigation is required to validate the bioactivity results.

## 6. Supplementary material

The following supplementary materials are available for this article:

**Figure S1.** GC/MS chromatogram of non-polar fraction of *A. palaestinum* spadices (EtOAc extract).

**Table S1.** % inhibition of HeLa cancer cell lines using different concentrations of the extracts.

**Table S2.** % inhibition of MCF-7 cancer cell lines using different concentrations of the extracts.

**Table S3.** % inhibition of HepG2 cancer cell lines using different concentrations of the extracts.

**Table S4.** Effect of *A. palaestinum* spadices crude extract towards membrane stabilization.

**Table S5.** Effect of *A. palaestinum* spadices/XAD extract towards membrane stabilization.

**Table S6.** Effect of *A. palaestinum* spathes crude extract towards membrane stabilization.

**Table S7.** Effect of acetylsalicylic acid towards membrane stabilization.

## References

- Aboul-Enein, A. M., El-Ela, F. A., Shalaby, E. A., & El-Shemy, H. A. (2012). Traditional medicinal plants research in Egypt: Studies of antioxidant and anticancer activities. *Journal of Medicinal Plants Research*, 6(5), 689–703. <https://doi.org/10.5897/jmpr11.968>
- Abu-Qatouseh, L., Mallah, E., & Mansour, K. (2019). Evaluation of anti-*Propionibacterium acnes* and anti-inflammatory effects of polyphenolic extracts of medicinal herbs in Jordan. *Biomedical and Pharmacology Journal*, 12(1), 211–217. <https://doi.org/10.13005/bpj/1629>
- Abu-Reidah, I. M., Ali-Shtayeh, M. S., Jamous, R. M., Arráez-Román, D., & Segura-Carretero, A. (2015). Comprehensive metabolite profiling of *Arum palaestinum* (Araceae) leaves by using liquid chromatography-tandem mass spectrometry. *Food Research International*, 70, 74–86. <https://doi.org/10.1016/j.foodres.2015.01.023>



- Azab, A. (2023). Top edible wild plants of Eastern Mediterranean region. Part I: Anticancer activity. *European Journal of Medicinal Plants*, 34(6), 29–51. <https://doi.org/10.9734/EJMP/2023/v34i61143>
- Booth, L., West, C., von Hoff, D., Kirkwood, J. M., & Dent, P. (2021). GZ17-6.02 interacts with [MEK1/2 and B-RAF inhibitors] to kill melanoma cells. *Frontiers in Oncology*, 11, Article 656453. <https://doi.org/10.3389/fonc.2021.656453>
- Dai, D. N., Huong, L. T., Hung, N. H., van Chinh, H., & Ogunwande, I. A. (2020). Antimicrobial activity and chemical constituents of essential oil from the leaves of *Alpinia globosa* and *Alpinia tonkinensis*. *Journal of Essential Oil-Bearing Plants*, 23(2), 322–330. <https://doi.org/10.1080/0972060X.2020.1752816>
- Devi, S., Ropiqa, M., Murti, Y. B., & Nugroho, A. K. (2020). Screening of extraction process and the estimation of total alkaloids in *Carica papaya* Linn. leaf. *Traditional Medicine Journal*, 25(2), 90–95. <https://doi.org/10.22146/mot.52184>
- El-Desouky, S. K., Hawas, U. W., & Kim, Y. K. (2014). Two new diketopiperazines from *Arum palaestinum*. *Chemistry of Natural Compounds*, 50(6), 1075–1078. <https://doi.org/10.1007/s10600-014-1162-y>
- El-Desouky, S. K., Ryu, S., & Kim, Y. (2007). Piperazirum, a novel bioactive alkaloid from *Arum palaestinum* Boiss. *Tetrahedron Letters*, 48(23), 4015–4017. <https://doi.org/10.1016/j.tetlet.2007.04.032>
- Farid, M. M., Hussein, S. R., Ibrahim, L. F., El Desouky, M. A., Elsayed, A. M., El Oqlah, A. A., & Saker, M. M. (2015). Cytotoxic activity and phytochemical analysis of *Arum palaestinum* Boiss. *Asian Pacific Journal of Tropical Biomedicine*, 5(11), 944–947. <https://doi.org/10.1016/j.apjtb.2015.07.019>
- Farid, M. M., Hussein, S. R., Trendafilova, A., Marzouk, M. M., El Oqlah, A. A., & Saker, M. M. (2017). Phytochemical constituents of the butanol fraction of *Arum palaestinum* Boiss: Cytotoxic and antiviral screening. *Journal of Materials and Environmental Science*, 8(7), 2585–2591.
- Halecki, W., Mozdzen, K., & Synowiec, A. (2022). Autoallelopathic and allelopathic influence of aqueous winter-cereal extracts. *Acta Agrobotanica*, 75(1), Article 753. <https://doi.org/10.5586/aa.753>
- Hamann, M. T. (2006). In S. D. Sarker, Z. Latif, & A. I. Gray (Eds.), *Methods in biotechnology. Natural products isolation* (2nd ed., p. 2666). Humana Press.
- Hatmal, M. M., Abderrahman, S. M., & Alsholi, D. (2017). Determining the anti-tumor effects of different extracting methods of *Arum palaestinum* on different cancer cell lines by *in vitro* assay. *Pharmacologyonline*, 1, 28–45.
- Hawash, M., Jaradat, N., Elaraj, J., Hamdan, A., Lebdeh, S. A., & Halawa, T. (2020). Evaluation of the hypoglycemic effect of seven wild folkloric edible plants from Palestine (Antidiabetic effect of seven plants from Palestine). *Journal of Complementary and Integrative Medicine*, 17(1), 1–10. <https://doi.org/10.1515/jcim-2019-0032>
- Ho, N. T. H., Thien, P. T., Nhat, Q. H., Hoang, A. N., Ngoc, T. L. T., Bao, L. N. D., Minh, N. N. T., & Bich, P. T. T. (2022). Microrhizome induction and curcumin accumulation in *Curcuma aromatica* Salisb. *Acta Agrobotanica*, 75(1), Article 7511. <https://doi.org/10.5586/aa.7511>
- Husein, A. I., Ali-Shtayeh, M. S., Jondi, W. J., Zatar, N. A. A., Abu-Reidah, I. M., & Jamous, R. M. (2014a). *In vitro* antioxidant and antitumor activities of six selected plants used in the Traditional Arabic Palestinian herbal medicine. *Pharmaceutical Biology*, 52(10), 1249–1255. <https://doi.org/10.3109/13880209.2014.886274>
- Husein, A. I., Ali-Shtayeh, M. S., Jamous, R. M., Zaitoun, S. Y. A., Jondi, W. J., & Zatar, N. A. A. (2014b). Antimicrobial activities of six plants used in Traditional Arabic Palestinian Herbal Medicine. *African Journal of Microbiology Research*, 8(38), 3501–3507. <https://doi.org/10.5897/AJMR2014.6921>
- Jaber, H. M., Al-Hamaideh, K. D., Al-Daghistani, H. I., Amer, N. H., Nassar, M. N., Saleh, M. H., Al-Latif, A., & Al-Nuaimi, A. H. (2020). Antibacterial activity and chemical composition of *Arum hygrophilum* Boiss crude extracts. *Jordan Journal of Biological Sciences*, 13(2), 159–164. <https://doi.org/10.13140/RG.2.2.30662.42562>
- Jaradat, N. A., & Abualhasan, M. (2016). Comparison of phytoconstituents, total phenol contents and free radical scavenging capacities between four *Arum* species from Jerusalem and Bethlehem. *Pharmaceutical Sciences*, 22(2), 120–125. <https://doi.org/10.15171/PS.2016.19>
- Jaradat, N. A., Al-Ramahi, R., Zaid, A. N., Ayesh, O. I., & Eid, A. M. (2016). Ethnopharmacological survey of herbal remedies used for treatment of various types of cancer and their methods of preparations in the West Bank-Palestine. *BMC Complementary and Alternative Medicine*, 16(1), Article 93. <https://doi.org/10.1186/s12906-016-1070-8>
- Kaur, R., & Arora, S. (2015). Alkaloids-important therapeutic secondary metabolites of plant origin. *Journal of Critical Reviews*, 2(3), 1–8.

- Khalaf, N. A., Naik, R. R., Shakya, A. K., Shalan, N., & Al-Othman, A. (2015). Antioxidant, anti-inflammatory and anti-diarrheal activity of ethanolic extract of some selected medicinal plants grown in Jordan and Palestine. *Oriental Journal of Chemistry*, 31(4), 1923–1928. <https://doi.org/10.13005/ojc/310408>
- Labbozzetta, M., Poma, P., Tutone, M., McCubrey, J. A., Sajeva, M., & Notarbartolo, M. (2022). Phytol and heptacosane are possible tools to overcome multidrug resistance in an *in vitro* model of acute myeloid leukemia. *Pharmaceuticals*, 15(3), Article 356. <https://doi.org/10.3390/ph15030356>
- Máthé, A., & Bandoni, A. (2021). *Medicinal and aromatic plants of the World*. Springer International Publishing. <https://doi.org/10.5860/choice.189230>
- Melariri, P., Campbell, W., Etusim, P., & Smith, P. (2012). *In vitro* and *in vivo* antimalarial activity of linolenic and linoleic acids and their methyl esters. *Advanced Studies in Biology*, 4(7), 333–349.
- Miah, M. M., Das, P., Ibrahim, Y., Shajib, M. S., & Rashid, M. A. (2018). *In vitro* antioxidant, antimicrobial, membrane stabilization and thrombolytic activities of *Dioscorea hispida* Dennst. *The European Journal of Integrative Medicine*, 19, 121–127. <https://doi.org/10.1016/j.eujim.2018.02.002>
- Naseef, H., Qadadha, H., Asfour, Y., Sabri, I., Al-Rimawi, F., Abu-Qatouseh, L., & Farraj, M. (2017). Anticancer, antibacterial, and antifungal activities of *Arum palaestinum* plant extracts. *World Journal of Pharmaceutical Research*, 6(5), 31–43. <https://doi.org/10.20959/wjpr201715-10091>
- Nisa, S., Bibi, Y., Masood, S., Ali, A., Alam, S., Sabir, M., Qayyum, A., Ahmed, W., Alharthi, S., Santali, E. Y., Alharthy, S. A., Bawazir, W. M., & Almashjary, M. N. (2022). Isolation, characterization and anticancer activity of two bioactive compounds from *Arisaema flavum* (Forssk.) Schott. *Molecules*, 27(22), Article 7932. <https://doi.org/10.3390/molecules27227932>
- Oliveira, A. L. S., Valente, D., Moreira, H. R., Pintado, M., & Costa, P. (2022). Effect of squalane-based emulsion on polyphenols skin penetration: *Ex vivo* skin study. *Colloids and Surfaces B: Biointerfaces*, 218, Article 112779. <https://doi.org/10.1016/j.colsurfb.2022.112779>
- Qnais, E., Bseiso, Y., Wedyan, M., & Alkhateeb, H. (2017). Evaluation of analgesic activity of the methanol extract from the leaves of *Arum palaestinum* in mice and rats. *Biomedical and Pharmacology Journal*, 10(3), 1159–1166. <https://doi.org/10.13005/bpj/1216>
- Rizvi, S. M. D., Shakil, S., Zeeshan, M., Khan, M. S., Shaikh, S., Biswas, D., Ahmad, A., & Kamal, M. A. (2014). An enzoinformatics study targeting polo-like kinases-1 enzyme: Comparative assessment of anticancer potential of compounds isolated from leaves of *Ageratum houstonianum*. *Pharmacognosy Magazine*, 10(1), 14–21. <https://doi.org/10.4103/0973-1296.127333>
- Shaaban, M. T., Ghaly, M. F., & Fahmi, S. M. (2021). Antibacterial activities of hexadecanoic acid methyl ester and green-synthesized silver nanoparticles against multidrug-resistant bacteria. *The Journal of Basic Microbiology*, 61(6), 557–568. <https://doi.org/10.1002/jobm.202100061>
- Shahriar, M., Ahsan, M. T., Hossain, S. M., Chowdhury, S. F., Mannan, M. M., Akhter, R., Mahmud, S., Hasan, S., & Narjish, S. N. (2015). *In vitro* membrane stabilizing activity of leaf extracts of *Paederia foetida* (L). *World Journal of Pharmaceutical Research*, 4(4), 204–209.
- Shi, M., Ho, K., Keating, A., & Shoichet, M. S. (2009). Doxorubicin-conjugated immuno-nanoparticles for intracellular anticancer drug delivery. *Advanced Functional Materials*, 19(11), 1689–1696. <https://doi.org/10.1002/adfm.200801271>
- Zargoun, H. M., Mohammed, M. A., Abdelshafeek, K. A., Ibansharrada, A. A., & Alomari, A. A. (2020). GC/MS analysis of lipid constituents and antimicrobial activity of *Arum cyrinaicum* extracts. *Egyptian Journal of Chemistry*, 63(12), 4889–4910. <https://doi.org/10.21608/EJCHEM.2020.30199.2647>